

Detection of antibodies of rinderpest and peste des petits ruminants viruses (*Paramyxoviridae*, *Morbillivirus*) during a new epizootic disease in Ethiopian camels (*Camelus dromedarius*)

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SUMMARY

A serological survey was designed to determine the antibody prevalence of rinderpest virus (RPV) and peste des petits ruminants virus (PPRV) in Ethiopian camels. That study was undertaken after the occurrence in 1995 of an apparently new highly contagious disease characterized by a rinderpest-like disease syndrome in the camel population. 90 dromedaries were distributed in groups based on three epidemiologically-defined regions. The first group was from a non-affected area, the second from sick and contact animals and the third from convalescent animals. The sera were analyzed for antibody to RPV and PPRV by competitive ELISA tests. Results showed a global seroprevalence of 7.8 % for PPRV antibodies and 21.3 % for RPV antibodies. None of the sera from the non-affected area was positive and the second and third groups had various positive rates. In accordance with several authors, the receptivity of the camel to these viruses appears to be a reality. However, its susceptibility to RPV and PPRV had never been confirmed, as well as its role as a potential reservoir of these viruses which cause two major diseases of ruminants. The hypotheses about the occurrence of an emerging infection in camels, caused by pathogens usually found in cattle, sheep and goats, are discussed.

KEY-WORDS : dromedary - serology - peste des petits ruminants - rinderpest - Ethiopia.

RÉSUMÉ

Détection d'anticorps de la peste bovine et de la peste des petits ruminants (*Paramyxoviridae*, *Morbillivirus*) lors d'une nouvelle maladie épi-zootique des dromadaires (*Camelus dromedarius*) observée en Ethiopie.
Par F. ROGER, M. GUEBRE YESUS, G. LIBEAU, A. DIALLO, L.M. YIGEZU et T. YILMA.

Un sondage sérologique a été réalisé afin de déterminer la présence d'anticorps dirigés contre les virus de la peste bovine (RPV) et de la peste des petits ruminants (PPRV) chez des dromadaires éthiopiens. Cette étude a été entreprise suite à la survenue en 1995 d'une maladie, apparemment nouvelle pour la population de dromadaires, hautement contagieuse et caractérisée par un syndrome de type peste bovine. 90 dromadaires ont été répartis en différents groupes de statuts épidémiologiques différents. Le premier groupe était constitué d'animaux d'une région non encore infectée, le second d'animaux malades et contacts et le troisième d'animaux convalescents. Les sérums ont été analysés par des tests ELISA de compétition. Les résultats ont montré globalement des taux de seroprévalence de 7.8 % pour les anticorps anti-PPRV et de 21.3 % pour les anticorps anti-RPV. Aucun des sérums du groupe non affecté n'était positif, alors que les second et troisième groupes montraient divers taux de séropositivité. Selon plusieurs auteurs, la réceptivité du dromadaire à ces virus est une réalité. Cependant, sa sensibilité n'a jamais été confirmée en Afrique, ainsi que son rôle comme un réservoir potentiel de ces virus responsables de maladies majeures des ruminants. Les hypothèses relatives à l'émergence d'une nouvelle infection chez le dromadaire, causées par des virus habituellement isolés chez les bovins, ovins et caprins, sont discutées.

MOTS-CLÉS : dromadaire - sérologie - peste des petits ruminants - peste bovine - Ethiopie.

Introduction

In the aim to provide elements of investigations after the occurrence of a rinderpest-like disease affecting from 1995 the camel population in Ethiopia, we designed a preliminary serological survey for the detection of antibodies of rinderpest virus (RPV) and peste des petits ruminants virus (PPRV). These diseases normally affect cattle and small ruminants and are caused by closely related viruses [9, 11].

The epizootic, previously described [21, 25], was characterized by a highly contagious respiratory syndrome with a

high rate of morbidity (over 90%) and a variable rate of mortality, which depended on the antibiotic treatment. All the camel population of Ethiopia, almost 2 millions of heads, had been affected in less than one year. Nomadic breeders of camels and the veterinary services had never encountered that type of disease.

The last outbreak of rinderpest was declared in 1994 in the North-East of Ethiopia and peste des petits ruminants is prevalent in Ethiopia for many years in sheep and goats [19, 23].

Material and methods

Sera collected from 90 dromedaries in three epidemiologically-defined regions were tested. The first group (G1) comprised 17 serum samples collected in August 1995 from Southern Ethiopia, an area considered non-affected by the disease, since the disease was confined to the northern part of the country. A second set (G2) was collected from 47 clinically sick or contact camels in Eastern Ethiopia in November 1995. A third group (G3) included 26 samples collected from convalescent camels in Northern Ethiopia one month after the outbreak ended in that region.

A competitive ELISA consisting of a monoclonal antibody to the PPR virus (PPRV) nucleoprotein (N) and recombinant PPRV N-protein as an antigen [14] was used for the quantification of PPRV antibodies in all 90 serum samples. A similar test was used for detecting RP virus (RPV) antibody that incorporated recombinant RPV hemagglutinin (H) protein and a specific anti-H monoclonal antibody [2] in 80 serum samples. The threshold value applied was 50 % of competition. Moreover, the quantitative values of competition were registered.

Results

PPRV serology (n = 90) - Serum samples from the non-affected area (G1) were all negative for PPRV antibodies, but 7 samples in groups G2 and G3 were tested positive for PPRV antibodies.

The qualitative values (percentage of positive) for the distinct groups were G1 = 0.0 %, G2 = 6.4 % and G3 = 15.4 % (graph 1).

The quantitative mean values calculated for each group on the basis of all samples are significantly different (table I).

RPV serology (n = 80) - Serum samples from the non-affected area (G1) were all negative for RPV antibodies, but 17 samples in group G2 and group G3 were tested positive for RP antibodies.

The qualitative (percentage of positive) values for the distinct groups were : G1 = 0 %, G2 = 36.4 % and G3 = 4.5 % (graph 1).

The quantitative mean values calculated for each group on the basis of all samples were significantly different (table II).

Discussion

Recent serological surveys have indicated the receptivity of the camels to RPV [3, 12, 18], although clinical signs have not been observed [6, 7, 8, 22, 24]. A study in Egypt showed 4.2 % of 142 healthy camels at a slaughterhouse were positive for PPRV antibodies by serum neutralization test [12]. However, a clinical disease in camels due to PPRV had not yet been established.

Considering the extremely rapid spread of the disease in Ethiopia, and in spite of the fact that the antibiotics had a positive effect, we can presuppose that the disease observed was initiated by a virus.

We hypothesize that the presence of PPRV and RPV antibodies in camels can be due :

- **either** to an immune humoral response to these viruses that expressed only the passage and an immune receptivity of the camel, without any clinical or sub-clinical expressions. In that case, we have to emphasize that the detection of these antibodies allow revealing the circulation of the RPV and PPRV among the cattle and small ruminants. Thus, in that hypothesis, the disease observed has been caused by other(s) pathogen(s).

- **or** to susceptibility to a morbillivirus closely related to RPV or PPRV, which was virulent for the camels. This virulence could be foreseen taking into account that morbilliviruses have an immunosuppressive effect [11] that led to secondary bacterial infections effectively treated by the antibiotics. Indeed, during this epizootic, *Streptococcus equi* sub-species *equi* have been isolated from sick camels in Afar region [25] and *Mannheimia haemolytica* from camels in Somali region [4]. Within the morbillivirus genus and related viruses, several emerging diseases have been recently described. The existence of mild strains of rinderpest in domestic cattle has had devastating effects in African buffaloes, eland and lesser kudu in Africa [1, 13, 17]. PPRV has been isolated during an outbreak in Indian buffaloes [10]. New morbillivirus with devastating effects have been described in marine mammals [15]. Canine distemper virus caused mortalities in lions of East Africa [20]. Viruses belonging to the *Paramyxoviridae* family, initially classified as morbillivirus, were recently described in diseases declared in horses and human [16] and pigs and human [5]. In that context, we presuppose that this new disease could result from an interspecies transfer of PPRV or RPV strains from cattle or goats and sheep to camel or through the emerging of a new morbillivirus, serologically related to PRV and/or RPV.

The first consequence of these results is that the camels could play a role in the epidemiology of these major diseases in Africa. Considering the necessary programs of eradication of rinderpest and control of peste des petits ruminants, camel populations should be integrated in the epidemiological surveys. They could especially be used for the epidemiosurveillance, during the process of the status evaluation of rinderpest and as sentinels of the disease.

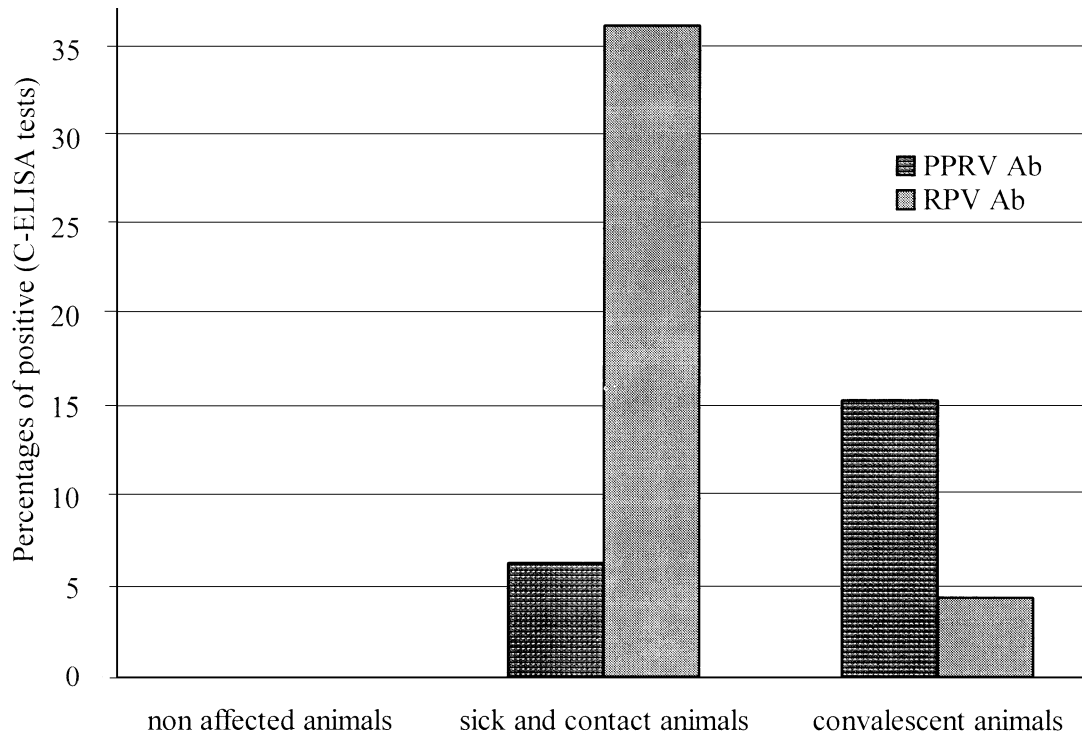
Microbiological, pathological and epidemiological investigations are in progress to precise the etiology of this apparently new disease of camels.

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GRAPH 1. — Qualitative values of PPRV and RPV antibodies.

Group	G1	G2	G3
Sample size	17	47	26
Mean for all sample *	0.247	0.323	0.391
Variance for all samples **	0.013	0.011	0.017
Minimum	0.000	0.095	0.195
Maximum	0.435	0.525	0.690
Mean for positive samples	-	0.513	0.619
Variance for positive samples	-	0.002	0.007

* ANOVA : $F = 8.14$; $p < 0.001$

** Variances are homogeneous with 95 % confidence (Bartlett's test for homogeneity)

TABLE I. — Values of competition among the three groups for PPR antibodies (N-protein).

Group	G1	G2	G3
Sample size	14	44	22
Mean for all sample *	0.229	0.399	0.243
Variance for all samples **	0.013	0.052	0.038
Minimum	0.113	0.032	0.000
Maximum	0.477	0.835	0.836
Mean for positive samples	-	0.658	0.836
Variance for positive samples	-	0.006	0.000

* Non parametric Kruskal-Wallis $H = 11.5$; $p < 0.01$

** Bartlett's test shows the variances in the samples differ (non parametric test is preferred rather than ANOVA)

TABLE II. — Values of competition among the three groups for RP antibodies (H-protein).

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